

Homo Sapien Bcr-Abl-Interacting Scaffolds from *Bryophyllum pinnatum* and *Cantharanthus roseus*: Computational Studies

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Abstract

Leukaemia is one of the ten life-threatening cancer of the world today. Chronic myelogenous leukaemia (CML), in particular constitutes about 15% of adult leukaemia and is characterised by over-production of immature myeloid cells in the bone marrow, spleen and peripheral blood resulting in the presence of the constitutively activated tyrosine kinase BCR-ABL, the cardinal therapeutic target. With the prevailing patients resistant to current drugs, it is imperative to come up with novel drugs particularly of plant origin considering the critical roles natural products (plants) play in the discovery and developments of new pharmaceuticals. In the present study, Virtual High Throughput Screening of phytochemicals from *Bryophyllum pinnatum* and *Cantharanthus roseus* (reported anticancer plants) against the Bcr-Abl catalytic site for possible drug scaffolds were carried out. A library of phytochemical compounds (ligands) was generated and multiple docking of the ligands against the Bcr-Abl catalytic site was executed with the Autodock Vina algorithm. Docking scores from Autodock vina were validated via the online available ChEMBL database and a correlation coefficient of 0.7 was obtained from a plot of the docking scores of 43 compounds from ChEMBL database with their corresponding pIC₅₀ values. The lead compounds from Virtual High Throughput Screening, Cynaroside and Rutin have a binding energies of -9.6 kcal/mol and -9.4 kcal/mol respectively when compared with the co-crystallized nilotinib's -9.4 kcal/mol. Pharmacological kinetics analysis indicated that Cynaroside, with a bit of optimization could end up a potential anti-CML drug.

Keywords: Leukemia, BCR-ABL Oncoprotein, *Bryophyllum pinnatum*, *Cantharanthus roseus*, Molecular Docking

Introduction

Leukaemia has a 94% death rate in Nigeria, only a paltry one out of every 20 Nigerians with leukaemia survives (1). A person with leukaemia has bone marrow that produces abnormal white blood cells called leukaemia cells and leukemic blast cells. Chronic myelogenous leukaemia (CML) is characterised by over-production of immature myeloid cells and mature granulocytes in the bone marrow, spleen and peripheral blood (2). Chronic myelogenous leukaemia (CML), in particular constitutes about 15% of adult leukaemia (3). The median age at presentation is 53 years, but all age groups, including children, are affected. The hallmark of CML is chromosomal translocation on t(9;22)(q34;q11), which explains the chimeric gene BCR-ABL formed on chromosome 22 (2). The effect of this gene is the ability to induce leukemic phenotype.

The BCR-ABL gene constitute an active kinase activity capable of activating and interfering with several cellular processes such as cell proliferation and differentiation leading to malignant transformation (4,5,6). Historically, it has been treated with chemotherapy, interferon and bone marrow transplantation, although targeted therapies introduced at the beginning of the 21st century have radically changed the management of CML. Insight into the cellular pathophysiology of CML has given rise to the development of various drug therapies. CML is treated with inhibitors of tyrosine kinase, the first with crystallographic material of which was imatinib mesylate (marketed as Gleevec or Glivec; previously known as STI-571). Among the various tyrosine kinase inhibitors developed is nilotinib, a second-generation inhibitor developed to be effective in imatinib-resistant patients. Imatinib resistance results from the emergence of point mutations that result from the kinase domain of BCR-ABL reducing the effectiveness of the drug (7, 8). Nilotinib is highly selective for wild-type BCR-ABL with 20 times the binding affinity of imatinib, the first drug developed towards CML (9).

Natural products play critical roles in the discovery and developments of new pharmaceuticals (10). Natural products have brought forth the most important success in the chemotherapy of cancer, most of the foremost anticancer drugs are unmodified natural products derived from plants or microorganisms (11). *Cantharanthus roseus* was reported to possess anti-cancer properties against leukemia (12). On the other hand, the anti-cancer

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properties of *Bryophyllum pinnatum* against leukaemia have not been properly defined, however when investigated with the human cervical cancer it was seen to show inhibitory properties (13). In the present study, Virtual High Throughput screening of phytochemicals from *Bryophyllum pinnatum* and *Cantharanthus roseus* against the target BCR-ABL catalytic site were carried out. The drug likeness and bioactivity of our hit compounds were determined through the calculation of their molecular properties; calculated logP, polar surface area, number of hydrogen bond donor, number of hydrogen bond acceptors and molecular weight (Lipinski's rule) (14).

Materials and Methods

Data Collection and Preparation

Phytochemicals characterised from *Cantharanthus roseus* and *Bryophyllum pinnatum* were obtained from literature. Fifty (50) phytochemicals each from *Cantharanthus roseus* and *Bryophyllum pinnatum* totalling one hundred phytochemicals from the two plants, were downloaded in sdf format from the PubChem database (<https://pubchem.ncbi.nlm.nih.gov>). The sdf format were converted to pdb format by Open Babel, and finally converted to pdbqt using ligprep command lines. The BCR-ABL oncoprotein structure with PDB ID: 3CS9 and crystallographic resolution of 2.50Å° was downloaded from the protein data bank (<http://www.rcsb.org>). The BCR-ABL oncoprotein in the crystal structure is in complex with the drug nilotinib (<http://www.rcsb.org>). PyMOL Autodock/Vina Plugin was employed to extract the receptor BCR-ABL and nilotinib. SWISS-MODEL software was used to rebuild missing residues and atoms of BCR-ABL (15).

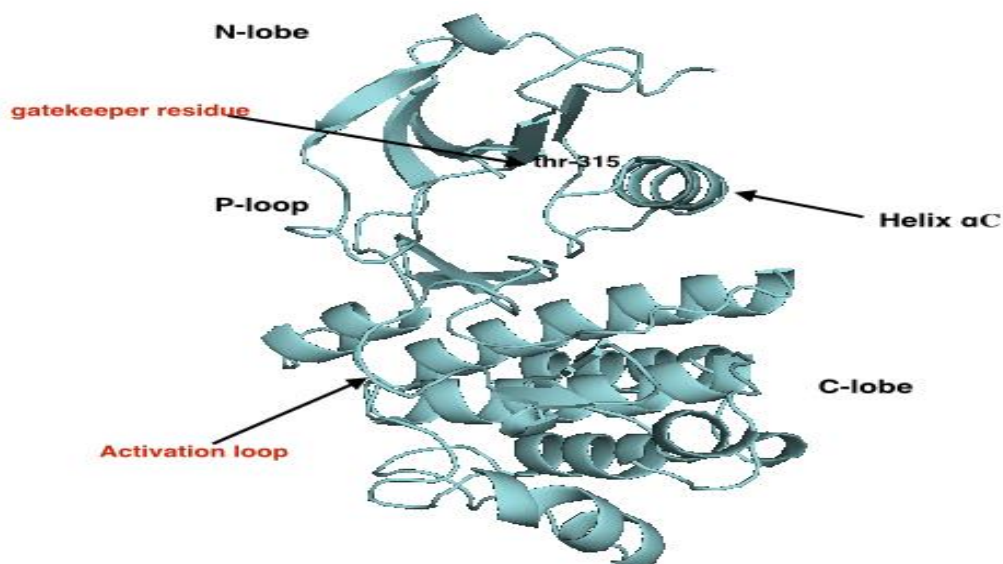


Figure 1: Structure of the BCR-ABL Oncoprotein

Virtual High Throughput Screening

A computational screening technique known as Virtual high throughput screening (vHTS), was used to screen a pool of compounds libraries to probe the binding affinity of the target receptor with the library compounds (16). The downloaded BCR-ABL from the protein data bank (<http://www.rcsb.org>), was uploaded in Pymol and a config file was generated, the grid coordinate was set as in the co-crystallized compound, x= -42.42, y= -50.7, z=-10.85. Our phytochemical compounds were converted to pdb, and to pdbqt, using command lines in auto dock vina and processed for docking.

Molecular Docking

The protein-ligand docking was carried out using Autodock Vina (17). The phytochemicals were docked into the exact BCR-ABL ATP catalytic site as occupied by the co-crystalized compound (nilotinib). Command lines in AutoDock Vina were used to carry out multiple docking of the phytochemicals.

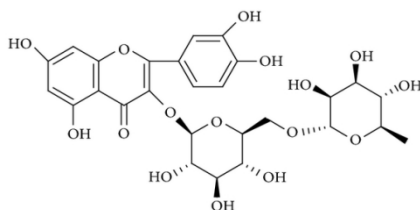
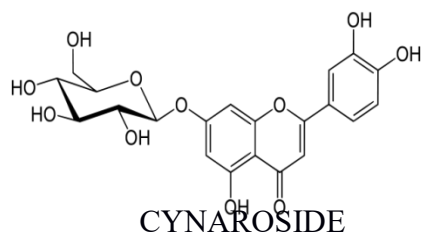
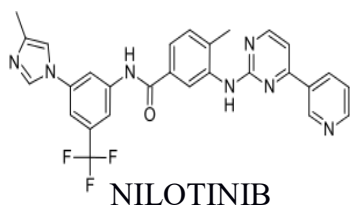
Validation of Docking Results

The Validation of result was performed with the multiple alignments of the BCR-ABL kinase domain receptor sequences obtained from pubmed. Using the online available ChemBL Database, the BCR-ABL sequences were blasted on www.ebi.ac.uk/chembl/. The result produced with the identity of 100%, IC₅₀ value of 1862 and KI value of 721, was downloaded in text format and converted to pdb format by Data Warrior version 2 (www.openmolecules.org), and finally converted to pdbqt. The 43 compounds obtained were docked into the Bcr-Abl catalytic site, as it was with the phytochemicals, being dictated by the config file (centre x=-42.42 ,

centre y=-50.7, centre z=-10.5, size x=22.50, size y=22.50, size z=22.50,). The config file indicates the grid map of the catalytic site of BCR-ABL kinase domain generated by AutoDock Vina. A correlation coefficient graph was plotted between the docking scores of the 43 compounds generated and their corresponding pIC₅₀ values.

Molecular Properties, Lipinski Rule “Rule Of Five”

According to Christopher Lipinski *et al.* (14), the rule of 5' predicts an orally active drug. Poor absorption or permeation is more likely when there are more than 5 H-bond donors, 10 H-bond acceptors, molecular weight (MWT) greater than 500 and the calculated Log P (CLogP) greater than 5 (or MlogP>4.15). The rule of five also describes the molecular properties, which are vital to ascertain the pharmacokinetics, which include the absorption, distribution, metabolism and excretion of compounds. We applied the Lipinski rule of five to determine the drug likeness and the probable biological activities of our lead ligands. The Marvin Viewer software (www.chemaxon.com) was employed to establish the conformity of our hit compounds to the rule of five. The number of rotatable bonds and polar surface area which are known to discriminate between compounds that are orally active and those that are not, for a large data set of compounds (18) were also derived. Compounds with 10 or fewer rotatable bonds as well as polar surface area equal or less than 140Å² have good oral bioavailability (18).



RUTIN

Figure 2: Chemical structure of Nilotinib and the lead compounds

Results and Discussion

Table 1: Docking scores of the phytochemicals from the anti-cancer plants (*Cantharanthus roseus* and *Bryophyllum pinnatum*). The two leads (cynaroside with the docking of -9.6 kcal/mol and rutin with the docking of -9.4kcal/mol) are from *Bryophyllum pinnatum*

<i>Cantharanthus roseus</i>		<i>Bryophyllum pinnatum</i>	
Phytochemicals	Docking Scores	Phytochemicals	Docking Scores
1-dodecanol	-4.8	24-epiclerosterol	-8
3-phenyldodecane	-6.1	alpa-amyrin	-5.9
ajmalicine	-8.8	alpha-amyrin acetate	-6.2
alpha-pinene	-5.1	astragalin	-8.2
alpha-terpineol	-5.6	bersaldegenin	-7.5
beta-bisabolene	-6.2	beta-amyrin	-5.4
beta-cadinene	-7.6	beta-amyrinacetate	-5.6
beta-phellandrene	-5.5	beta-sorbitol	-8.6
beta-pinene	-5	bufa-20,22-dienolide	-6.1
camphor	-4.9	campesterol	-8.8
catharathine	-8	chamomile	-8.3
cedrol	-7.8	citric acid	-4.4
davanone	-6.6	clerosterol	-7.1
dehydroabietic acid	-7.7	cynaroside	-9.6
delta-cadinene	-6.2	cysteine	-3.2
dodecane	-4.8	dimethylcaffeic acid	-6.2
dodecyl acetate	-5.2	d-sorbitol	-4
farnesyl acetone	-6.4	ferulic acid	-6.4
gamma-teripene	-4.6	friedelin	-7.6
geranial	-5.4	glucose	-4.6
geraniol	-5.4	glutamic acid	-4.3
hexadecane	-5.1	glycine	-3
iso-dihydrocarveol	-5.4	isocitric acid	-4.6
isopulegol	-5.5	isoquercitrin	-8.2
lanosterol	-7	malic acid	-4.1
limonene	-5.4	methione	-3.8
linalool	-5.4	nicotinic acid	-4.6
methyleugenol	-5.5	oxalic acid	-3.5
neral	-5.4	palmitic acid	-5.4
pentadecane	-5.2	patuletin	-8.2
perillaldehyde	-5.6	p-coumaric acid	-6.2
phytol	-6.1	pepostanol	-7.7
reserpine	-7.4	phenylalanine	-5.7
roridin D	-7.9	psi-taraxerol	-8
tabersonine	-7.7	pyridoxine	-5
terpinen-4-ol	-5.3	quercitrin	-8.4
tetracontane	-6.1	raffinose	-6.6
tetradecanal	-5	riboflavin	-7.8
trans-alpha-bergamot	6.7	rutin	-9.4
trans-sabienene hydr	5.1	salicylic acid	-5.1
tridecane	-4.7	soyasaponin	-7.3
trimethyldodecane	-5.9	stearic acid	-5.6
vanillic acid	-5.4	stigmasterol	-7.7
vinblastine	-5.7	succinic acid	-4
vincamine	-8.5	syringic acid	-5.6
vincristine	-7.1	taraxerol	-5.6
vindesin	-6.3	thiamine	-6.3
vitamin E	7.7	tyrosine	-6.1
widdrol	-5.3	vanillic acid	-5.4
(Z)-beta-ocimene	-5.4	vit.E acetate	-7.1

Table 2. Molecular properties of the lead compounds (cynaroside and rutin) and the standard drug (nilotinib)

S/N	Compounds	M w	HBD	HBA	ClogP	RTY	RB	PSA
1	Nilotinib (standard Drug)	529.53	2	9	4.9	152.85	6	97.62
2	Cynaroside	448.38	7	11	0.5	107.04	4	186.37
3	Rutin	610.52	10	16	-1.3	140.15	6	265.52

M w= Molecular Weight

HBD= Hydrogen Bond Donor

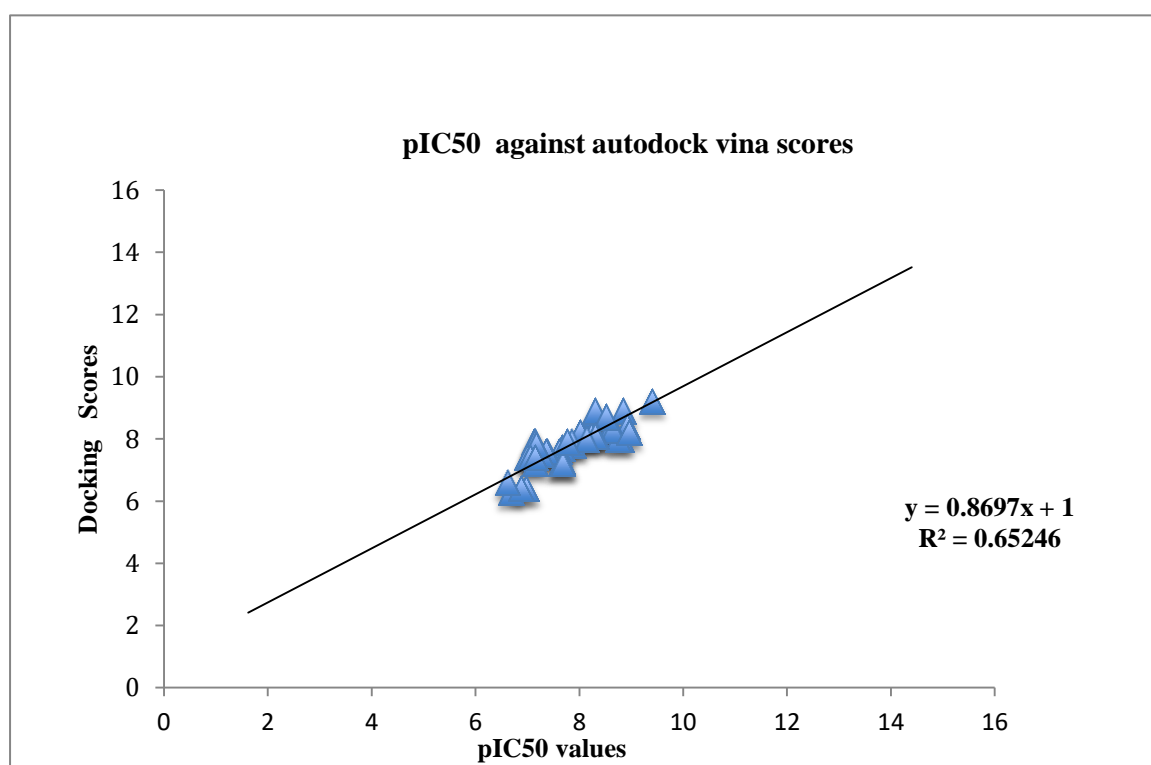
HBA= Hydrogen Bond Acceptor

ClogP= Calculated Log P(lipophilicity)

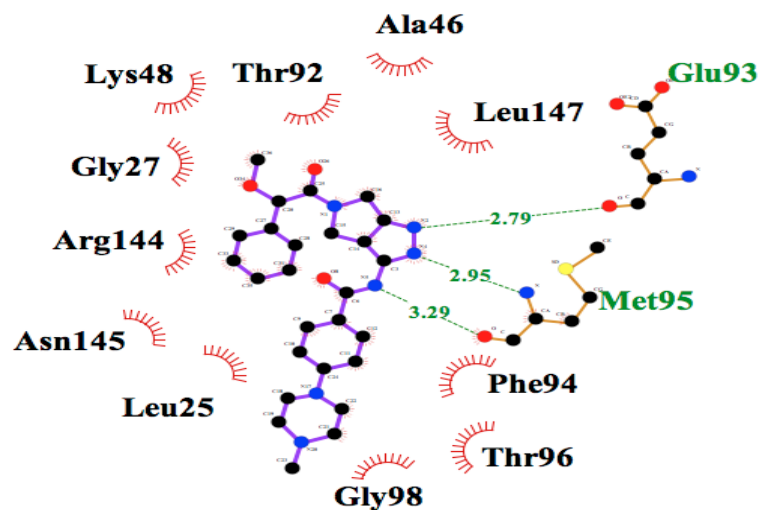
RTY= Refractivity

RB= Rotatable Bond

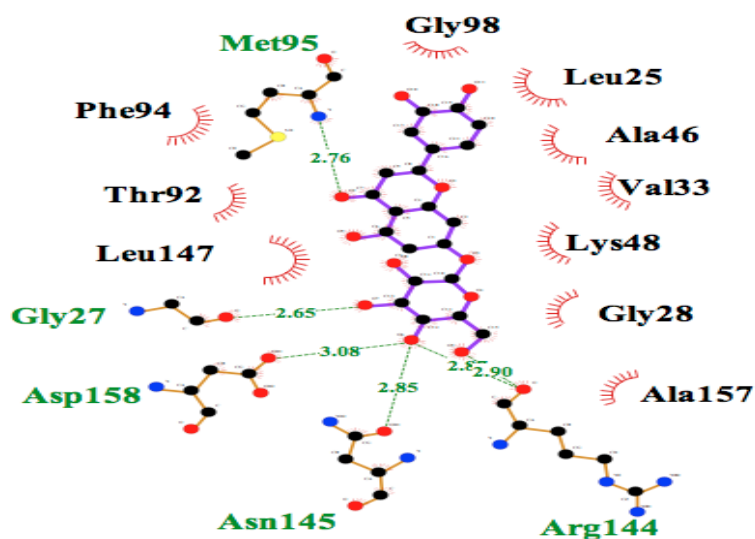
PSA= Polar Surface Area.

Figure 4: Autodock vina Scores vs Experimental pIC₅₀ values for 43 compounds docked into the Bcr-Abl catalytic site.

a).



b).



c).

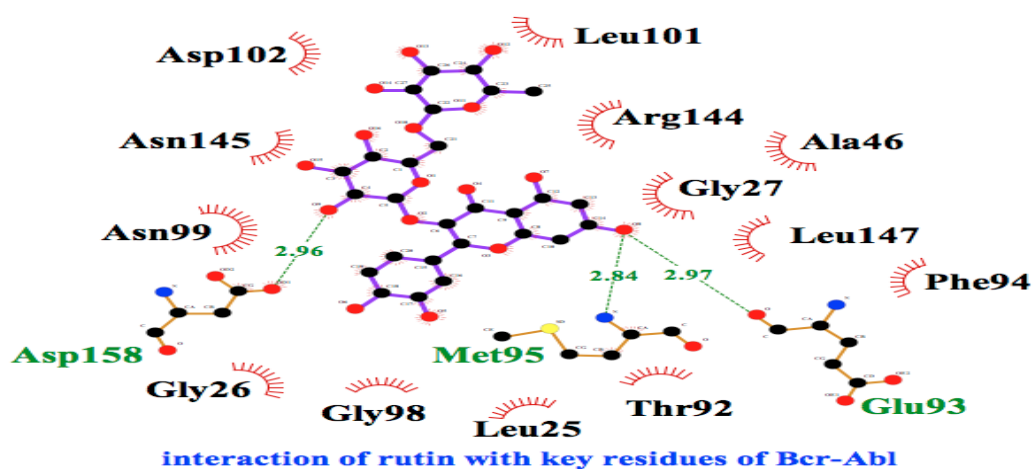


Figure 3: Interactions of nilotinib and the hit compounds with key residues of the therapeutic target, Bcr-Abl

Chronic myeloid leukaemia (CML) is a haematological cancer of the white cell, that contributes about 15% of adult leukaemia (19). With the increased cases of chronic myeloid leukaemia associated death (19), there is need for more potent drugs especially of plants origin; plants have brought forth the most important success in the chemotherapy of cancer (11). In this present study phytochemicals from *Cantharanthus roseus* and *Bryophyllum pinnatum* were docked into the catalytic site of Bcr-Abl using nilotinib, an FDA approved drug as the standard. From our virtual highthrough-put screening (vHTS) it is worthy of note that only *bryophyllum pinnatum* has phytochemicals with lower binding energy (Cynaroside with -9.6kcal/mol and rutin with -9.4kcal/mol) when screened against the Bcr-Abl catalytic site (Table 1), though *Cantharanthus roseus* has been reported to have anti-leukemic properties (12).

Cynaroside, with binding energy -9.6kcal/mol and rutin with -9.4kcal/mol were found to have a better inhibitory effect on Bcr-Abl catalytic site when compared with the co-crystallized nilotinib (-9.2kcal/mol). The closely related binding energies between cynaroside, rutin and the standard nilotinib, is believed to be as a result of the hydrogen bond interactions with key residues. Cynaroside forms six hydrogen bond interactions with ASP 158, ASN 145, ARG 144, GLY 27 (Figure 3 b), rutin forms three hydrogen bond interactions with GLU 93, MET 95, ASP 158, while nilotinib forms three hydrogen bonds with GLU 93 and MET 95 (Figure 3 c). Cynaroside, rutin and the standard drug nilotinib all share a common hydrogen bond interaction with MET 95, which explains the closely related binding affinities.

Cynaroside with a better binding affinity is probably due to the greater number of hydrogen bond interactions (6H-bond) when compared with rutin and nilotinib with three hydrogen bond each (3H-bond) (Figure 3). Our lead compounds were subjected to lipinski's rule of five (14) to evaluate their possible oral bioavailability and pharmacokinetics. It was shown that cynaroside obeyed the rule with a molecular weight of 448.38 and ClogP value of 0.5 and a refractivity of 107.04 and rotatable bond of 4 while rutin violated the lipinski's rule. It is worthy of note that both cynaroside and rutin have been reported elsewhere to possess anti-cancer effects (20, 21). Both cynaroside and rutin are potential anti-leukaemia phyto-compounds. A positive correlation of 0.67 (Figure 4) shows that our docking scores using AutoDock Vina algorithm is dependable and depict possible experimental pIC₅₀ values obtainable.

Conclusion

Virtual High Throughput screening of Phytochemicals from *Cantharanthus roseus* and *Bryophyllum pinnatum* against the Bcr-Abl catalytic site, revealed rutin and cynaroside from *Bryophyllum pinnatum* as the hits compounds. Both cynaroside and rutin are potential anti-leukaemia phyto-compounds.

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References

1. Nzelu A: Call to Action for Leukaemia (Blood Cancer). *The Guardian*, Retrieved from <https://guardian.ng/opinion/call-to-action-for-leukaemia-blood-cancer/>. 2015, September 19.
2. Thompson PA, Kantarjian H, Cortes JE: Diagnosis and Treatment of Chronic Myeloid Leukemia (CML) in 2015. *Mayo Clinic Proceedings*, 90(10): 1440- 14. <http://doi.org/10.1016/j.mcp.2015.08.014>. 2015.
3. Updyke KM, Morales-Lappot J, Lee T: Atypical Presentation of Chronic Myelogenous Leukemia. *Cureus*, 9(5),e1280.<http://doi.org/10.7759/cureus.1280>. 2017
4. Daley GQ, Baltimore D: *Proceedings of the National Academy of Sciences of the United States of America* 85, 9312 – 9316. 1988.
5. Daley GQ, Van Etten RA, Baltimore D: *Science*, 247, 824 – 830. (1990).
6. Gishizky ML, Johnson-White J, Witte ON: *Proceedings of the National Academy of Sciences of the United States of America* 90, 3755 – 3759. 1993.
7. Cowan-Jacob SW, Guez VM: (STI571) resistance in chronic myelogenous leukemia: molecular basis of the underlying mechanisms and potential strategies for treatment." *Mini Review in Medicinal Chemistry* 4(3): 285-99. 2004
8. Hochhaus A, La Rosee P: "Imatinib therapy in chronic myelogenous leukemia: strategies to avoid and overcome resistance. *Leukemia* 18:1321-31. 2004.
9. Blay JY, von Mehren M: Nilotinib: a novel, selective tyrosine kinase inhibitor. *Seminars in oncology*, 38 Suppl 1(0 1), S3-9. 2011.
10. Socjarto OO, Farnsworth NR: Bioactive Compounds from Natural Sources. *Perspective in Biology and Medicine*, 32: 244. 1998.

11. Loo TL, Freireich EJ: Cancer Chemotherapeutical Basic Concepts and Clinical Applications. *Chapman and Hall* 1475-1516. 1995.
12. Moteriya P, Rinkal S, Chanda S: Screening of phytochemical constituents in some ornamental flowers of Saurashtra region. *Journal of Pharmacognosy and Phytochemistry*, **3**(5): 112-120. 2014.
13. Mahata S, Maru S, Shukia S, Pandey A, Mugesh G, Das BC *et al*: Anticancer property of *Bryophyllum pinnata* (Lam.) Oken leaf of human cervical cancer cells. *BMC Complementary and Alternative Medicine* 12-15. 2012.
14. Lipinski CA, Lombardo F, Dominy BW, Feeney PJ: Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Advanced Drug Delivery Reviews*, **46**: 3-26. 2001.
15. Bordoli L, Kiefer F, Arnold K., Benkert P, Battey J, Schwede T: Protein structure homology modelling using Swiss-MODEL workspace. *National protocol* **4**(1): 1-13. 2009.
16. Shoichet KB: Virtual screening of chemical libraries, *Nature* **432**: 862–865 doi: 10.1038/nature03197. 2004.
17. Trott O, Olson AJ AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization and multithreading. *Journal of Computational Chemistry*, **31**(2):455-461. 2010. <http://doi.org/10.1002/jcc.21334>
18. Veber DF, Johnson SR, Cheng HY, Smith BR, Ward KW, Kopple KD: Molecular properties that influence theoral bioavailability of drug candidate. *Journal of Medicinal Chemistry* **5**(12):2615-23. 2002.
19. O'Dwyer ME, Mauro MJ, Kurilik G, Mori M, Balleisen S, Olson S *et al*: The impact of clonal evolution on response to imatinib mesylate (ST1571) in accelerated phase CML. *Blood*, **100**:1628-1633. 2002.
20. Chen H, Miao Q, Geng M, Liu J, Hu Y, Tian L, Yang Y: Anti-Tumor Effect of Rutin on Human Neuroblastoma Cell Lines through Inducing G2/M Cell Cycle Arrest and Promoting Apoptosis. *The Scientific World Journal*, 269165. <http://doi.org/10.1155/2013/269165>. 2013.
21. Thuy BTP, Nhung NTA, Duong T, Trung PV, Quang NM, Dung HTK *et al*: Prediction of anticancer activities of cynaroside and quercetin in leaf of plants *Cynara scolymus* L and *Artocarpus incisa* L using structure–activity relationship. *Cogent Chemistry*,2(1),1212452.<https://doi.org/10.1080/23312009.2016.1212452>. 2016