

Anti-Obesity Potential of the Ethanolic Extract of *Alstonia boonei* Stem Bark on High Carbohydrate Diet Induced Obesity in Male Wistar Rats

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ABSTRACT: Obesity increases the likelihood of various diseases, particularly dyslipidemia, type 2 diabetes, hypertension, heart disease and osteoarthritis. The aim of this study was to determine the anti-obesity potential of the ethanolic extract of *Alstonia boonei* stem bark on diet induced obesity (DIO) in male Wistar rats. Thirty male Wistar rats were divided into three groups (n=10): Animals in group 1 were fed normal control diet, Group 2 and 3 rats were fed high carbohydrate diet for 14 weeks, which was the period of obesity induction. The treatment period lasted for the next 4 weeks in which the rats in Group 1 and 2 received the vehicle (normal saline) and only Group 3 received 500 mg/kg b.w *Alstonia boonei* extract. The results obtained showed that feeding rats with *Alstonia boonei* extract significantly decreased ($p < 0.05$) food intake, body weight, fat mass, adiposity index, TC, LDL-C, leptin levels. It however had no significant effect on HDL-C, glucose and insulin levels when compared with the HCD obese control. The photomicrograph of HCD obese controls showed presence of numerous prominent fat deposits in the livers; this appeared reduced among the groups treated with the plant extract. Thus, the ethanolic extract of *Alstonia boonei* stem bark may have some potential in the management and/or treatment of obesity.

Key words: Obesity, *Alstonia boonei*, Fat mass, Adiposity index and Diet.

Introduction

Obesity is reaching epidemic proportions worldwide; it is correlated with various comorbidities, among which the most relevant are dyslipidemia¹, diabetes mellitus type 2², fatty liver³, and cardiovascular diseases such as heart failure and coronary heart disease⁴. As of 2005 the WHO estimated that at least 400 million adults (9.8%) are obese, with higher rates among women than men¹. Almost 70% of adults in U.S.A. are overweight but, perhaps more alarmingly, 16% of juveniles are overweight⁵.

Overweight and obesity are often defined simply as a condition of abnormal or excessive fat accumulation in adipose tissue⁶. While genetic influences are important to understanding obesity, they cannot explain the current dramatic increase seen within specific countries or globally⁷. According to Slyper⁸, in the United States, simple carbohydrate intake may be blamed for the rise in obesity, particularly in children who regularly consume fruit juices and snacks like potato chips. Foods high in fat or sugar (for example, fast food, fried food, and sweets) have high energy density. Carbohydrates increase blood glucose levels, which in turn stimulate insulin release by the pancreas, and insulin promotes the growth of fat tissue and can cause weight gain. Some scientists believe that simple carbohydrates (sugars, fructose, desserts, soft drinks, beer, wine, etc.) contribute to weight gain because they are more rapidly absorbed into the blood-stream than complex carbohydrates (pasta, brown rice, grains, vegetables, raw fruits, etc.) and thus cause a more pronounced insulin release after meals than complex carbohydrates. This higher insulin release is believed to contribute to weight gain.

It's easier to prevent obesity than to cure it. But in the event of obesity, its management and treatment includes dieting, exercising, surgery and medications such as Sibutramine and Orlistat. These drugs are more effective in people who make dietary changes and exercise more than in people who just take the drugs. As with almost all medications, weight-loss drugs have side effects, high cost and are not safe for everyone.

Traditional medicinal plants are often cheaper, locally available, and easily consumable (raw or as simple medicinal preparations). Some traditional healers have claimed that some medicinal plants in Nigeria, like *Alstonia boonei* could be used to treat obesity. The common name of *Alstonia* in English is *Alstonia*, cheesewood, timber tree — pattern wood, stoolwood and French: emien. In Nigeria, it is called égbū or *egbun* (Igbo), ahùn (Yoruba), Ukhu (Edo) and Ukpukunu (Urhobo). *A. boonei* De Wild is a tree of 30–40 m high by over 3 m in girth, bole cylindrical and long to 27 m, with high narrow buttresses. *Alstonia boonei* De Wild is a medicinal plant used extensively in West and Central Africa for the treatment of malaria, fever, intestinal helminthes, rheumatism, hypertension, etc^{9, 10}. This study is aimed to ascertain the anti-obesity potential of the ethanolic extract of *Alstonia boonei* De Wild stem bark on high carbohydrate diet induced obesity in male Wistar rats.

Materials and Methods

Plant Material: The fresh stem bark of *Alstonia boonei* De Wild was collected from a farm land located in Umuekwune community, Imo State, Nigeria. The authentication of the plants was done by a taxonomist at the Forestry Research Institute of Nigeria (FRIN), Ibadan, Oyo State. A voucher specimen of the plant was deposited in the Herbarium, FRIN.

Preparation of Plant Extract: The fresh parts of the stem bark of *A. boonei* was washed, chopped into pieces and air-dried in the laboratory at room temperature. The dried plant part was milled into powder and weighed. 250g of the plant powder was soaked in 500 ml of absolute ethanol in bottles for 72 hours with intermittent shaking. Then, it was filtered through Whatman No. 1 filter paper. The resulting filtrates were evaporated under reduced pressure using a rotary evaporator and there after freeze dried to a powder. The yield was stored in a refrigerator (4 °C) until required.

Phytochemical Screening: Phytochemical screening of the stem bark of *A. boonei* De Wild was done according to methods described by Treatise and Evans¹¹ and Sofowora¹².

Acute Toxicity (LD₅₀) of Plants: The acute toxicity of the ethanolic extract of the plant was carried out as described by Shah *et. al.*¹³ and Burger *et. al.*¹⁴.

Composition of Experimental Diet:

The food/supplement used to compose the diet include: carbohydrate (processed cassava locally called *garri*), protein (dried *bongafish*), fat (butter), fibre, vitamins and mineral mixture were obtained from a retail goods manufactured by SuperMax Nig. Ltd (Table 1).

Table 1: Composition of Experimental Diet

C o m p o s i t i o n	F o o d / S u p p l e m e n t s	N P D (%)	H C D (%)
C a r b o h y d r a t e s	G a r r i	6 0 . 0	6 0 . 0
	S u c r o s e	-	2 0 . 0
F a t	B u t t e r	5 . 0	5 . 0
P r o t e i n s	B o n g a F i s h	3 0 . 0	1 0 . 0
F i b e r		1 . 5	1 . 5
M i n e r a l m i x t u r e	M u l t i - m i n e r a l s	2 . 5	2 . 5
V i t a m i n m i x t u r e	M u l t i - v i t a m i n s	1 . 0	1 . 0
E n e r g y (K C a l / g)		4 . 0 9 5	4 . 0 9 5

Note: NPD-normal pellet diet; HCD- high carbohydrate diet

Experimental Design and Animal Grouping: Thirty male Wistar rats (average weight 80-90 g) were divided into 3 groups containing 10 rats per group. Group 1 was fed on normal pelleted diet, while obesity was induced in Groups 2 and 3 by feeding high carbohydrate diet (HCD) for 14 weeks (Table 1). Thereafter, only the rats in group 3 were treated with 500mg/kg b.w *Alstonia boonei* extract for the next 4 weeks. The rats had free access to their diets and water.

Food Intake and Body Weight Measurement: Daily food intake and weekly body weight was measured in grams (g).

Adipose Tissue Dissection and Fat Mass Determination: After abdominal incision, five different white adipose depots (two subcutaneous and three intra-abdominal) and interscapular brown adipose tissue were harvested from each rat and referred to as fat mass.

Adiposity Index: Adiposity index was determined by the sum of epididymal, visceral and retroperitoneal fat weights divided by body weight × 100, and expressed as adiposity percentage¹⁵.

Blood Sample Preparation and Bioassays: At the end of the experiment, rats were fasted for 12 to 14 hours. Blood was collected by cardiac puncture into plain sample bottles, allowed to stand for 15mins for the blood to retract, and then centrifuged at 3000 rpm for 5mins. The serum was separated and used for the assay of the following parameters: total cholesterol¹⁶, triglycerides¹⁷, high density lipoprotein cholesterol¹⁸, very low density lipoprotein cholesterol (triglycerides/5), low density lipoprotein cholesterol¹⁹, glucose¹⁶, insulin (DRG Insulin ELISA kit, USA) and leptin levels (DRG Leptin ELISA kit, USA).

Histopathology

The parts of the liver and heart were dissected out and placed in formal saline for histopathology studies. Sections of the liver and heart were prepared and stained with haematoxylin and eosin following fixation. Permanent mounts were examined by light microscopy and the results obtained were compared with control²⁰.

Statistical Analysis: The experimental results was expressed as the Mean ± S.E.M. Statistical significance of difference in parameters amongst groups was determined by One way ANOVA followed by Duncan's multiple range test. P<0.05 was considered to be significant.

Results

The bioactive compounds in the ethanolic stem bark extract of *Alstonia boonei* include alkaloids, saponins, tannins, steroids, flavonoids, cardiac glycosides and terpenoids. The acute toxicity (LD₅₀) study of *A.boonei* revealed general weakness and sluggishness as the major behavioral changes observed in the rats at 6400 mg/kg b.w oral dose. These behavioural changes disappeared after 1 hour of observation. No death was recorded at any of the doses administered. Oral LD₅₀ was therefore not determined because mortality was not observed.

The food intake, body weight, fat mass and adiposity index of the HCD obese control was increased significantly (P<0.05) compared to the normal control at the 18th week. The group treated with *A.boonei* extract had significant decrease (P<0.05) in the food intake, body weight, fat mass and adiposity index compared to the obese control (Table 2).

Table 2: Food intake, body weight, fat mass and adiposity index of rats treated with *A. boonei* extract.

G R O U P	Food Intake (g)	Body Weight (g)	Total Fat Mass (g)	Adiposity Index
N o r m a l C o n t r o l	63.13 ± 2.11 ^b	293.67 ± 9.17 ^c	20.58 ± 0.50 ^c	3.29 ± 0.18 ^b
H C D O b e s e C o n t r o l	76.72 ± 1.64 ^a	399.33 ± 7.06 ^a	29.43 ± 0.68 ^a	4.63 ± 0.11 ^a
H C D + A . b o o n e i	40.66 ± 2.73 ^c	357.00 ± 7.37 ^b	22.67 ± 0.72 ^b	3.27 ± 0.20 ^b

Values are expressed as means ± SEM. Means in the same column not sharing common letter(s) are significantly different (p < 0.05).

Table 3 shows the lipid profile of rats treated with *A.boonei* extracts. The HCD obese rats had significantly increased (P<0.05) serum triglycerides, total cholesterol, VLDL cholesterol and LDL cholesterol, with a decrease in HDL cholesterol which was not significant (P<0.05). All the parameters with exception of LDL-C of the treated group 3 had no significant decrease when compared to HCD obese control.

Table 3: Lipid profile of rats treated with the *A. boonei* extract

G R O U P	TG (mg/dl)	TC (mg/dl)	HDL-C(mg/dl)	VLDL-C(mg/dl)	LDL-C (mg/dl)
N o r m a l C o n t r o l	45.58 ± 0.91 ^b	43.78 ± 1.45 ^c	29.97 ± 1.70 ^c	9.12 ± 0.11 ^b	4.68 ± 0.50 ^b
H C D O b e s e C o n t r o l	83.57 ± 5.00 ^a	54.50 ± 2.09 ^{ab}	23.98 ± 1.51 ^{ab}	16.71 ± 1.00 ^a	13.82 ± 1.39 ^a
H C D + A . b o o n e i	83.87 ± 3.56 ^a	46.39 ± 4.83 ^b	22.48 ± 3.81 ^b	16.77 ± 0.63 ^a	7.13 ± 0.7 ^b

Values are expressed as means ± SEM. Means in the same column not sharing common letter(s) are significantly different (p < 0.05).

Serum glucose and insulin concentrations were significantly higher (P<0.05) in HCD obese control, while no significant increase (P<0.05) was observed in leptin level when compared to the normal control. On the other hand, the HCD obese rats treated with *A. boonei* had no significant

difference ($P < 0.05$) in glucose and insulin concentration, but showed significant decrease ($P < 0.05$) in leptin compared to the normal and HCD obese control (Table 4).

Table 4: Blood glucose, insulin and leptin of rats treated with the *A. boonei* extract

Table 4: Blood glucose, insulin and leptin in rats treated with the A. boonei extract															
G		R	O	U	P	GLUCOSE (mg/dl)		INSULIN (μ lU/ml)		LEPTIN (ng/ml)					
Normal Control						65.000	±	2.89 ^b	±	2.33	±	0.03 ^b	2.63	±	0.15 ^b
HCD Obese Control						89.67	±	3.18 ^a	±	2.50	±	0.06 ^a	2.86	±	0.09 ^{a b}
HCD + A. boonei						89.67	±	6.06 ^a	±	2.40	±	0.00 ^{a b}	0.80	±	0.12 ^c

Values are expressed as means $\pm$ SEM. Means in the same column not sharing common letter(s) are significantly different ($p < 0.05$).

In the histopathology of the liver, the cells of normal control (A) were compact; hepatocytes appeared distinct with relatively few tiny fats deposits scattered around the sinusoidal space and the central vein appeared distinct. The HCD obese control (B) had some fat deposits scattered around the sinusoidal space. Also, there was a little distortion in the architecture of the cells. The liver of *A. boonei* treated rats looked coarse as though a reaction was taking place and the fat deposits appeared reduced compared to the HCD obese control. In the histopathology of the heart, the cardiac muscle fibre of normal control (A) showed the myocardium separated by intercalated disc with no fat deposits. Similarly, no fat deposits were seen in the HCD obese and treated groups although there were signs of mild myocardial infarction.

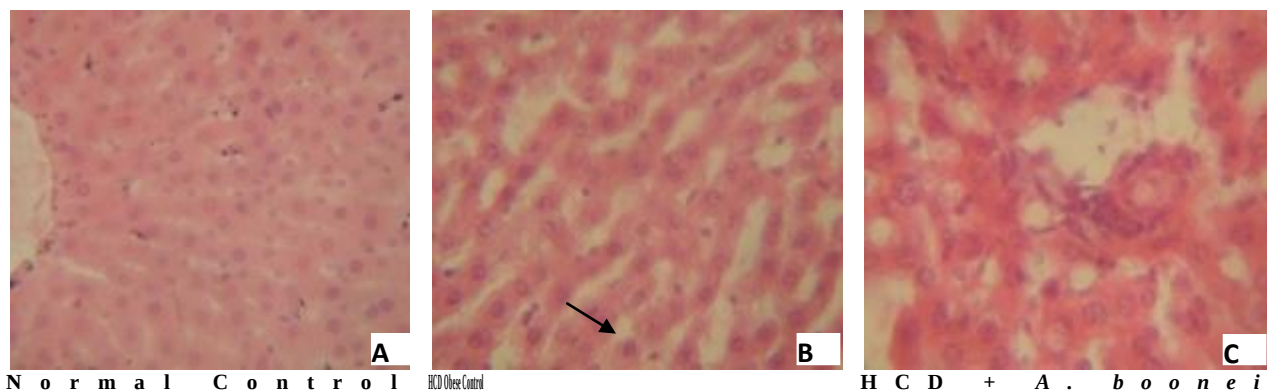


Plate 1: Sections of liver of normal control (A) without fat deposits, HCD obese control (B) with fat deposits and treated animals (C) showed reduced fat deposits (X40).

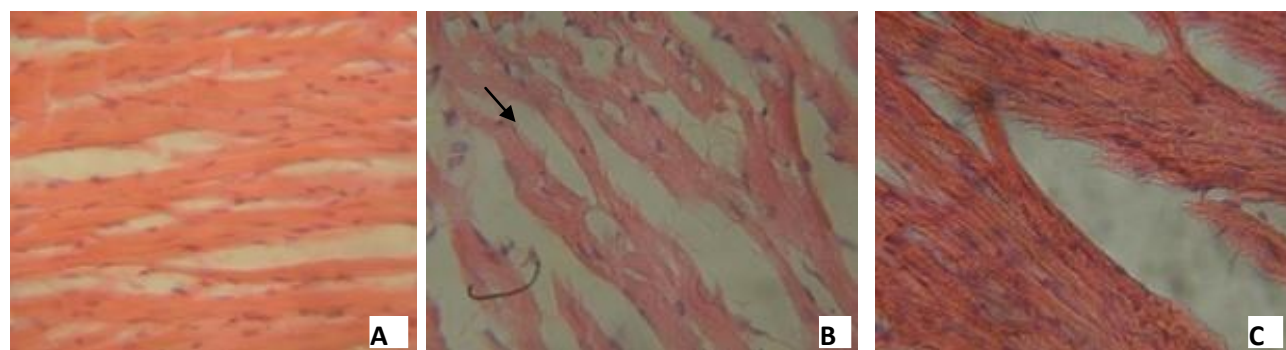


Plate 2: Sections of heart of normal control (A), HCD obese control and treated animals (C) showed no fat deposits (X40).

Discussion and Conclusion

The phytochemical screening of the ethanolic extract of *Alstonia boonei* stem bark shows the presence of alkaloids, saponins, tannins, steroids and flavonoids. This is consistent with the earlier reports of Akinmoladun *et al.*²¹, Amole and Ilori²² and Onyeneke and Anyanwu²³.

The increase in food intake in the HCD groups may be due to the palatability of the diet as refined sugar (sucrose) was part of its ingredients and sucrose is a combination of glucose and fructose. Sugar metabolism occurs primarily in the liver, where a high fructose flux leads to enhanced accumulation of hepatic triglycerides, resulting in impaired glucose and lipid metabolism²⁴, resulting in a high level of fat in storage. The consequence of high fat storage is body weight gain or increase, and then obesity. In the present study, obesity from the continuous increase in body weight as a consequence of the HCD. Similar results had earlier been obtained in body weight changes with the feeding of a high carbohydrate diet^{25, 26, 27, 28} and adiposity index²⁹.

Alstonia boonei caused significant decrease in food intake in HCD obese rats. This suggests that the plant might have appetite suppressing ability. The observed decrease in food intake would be accompanied with lower exposure to calorie intake which would have affected the body weight of the group treated with *A. Boonei*. In a toxicity study, it had been reported that *A.boonei* decreased significantly the body weights of rats treated for 4 and 12 weeks³⁰.

The significantly higher fat mass and adiposity index of HCD obese control when compared to the normal control rats corroborates the results found in other studies³¹. However, *A. boonei* extract produced a significant decrease in the total fat mass and adiposity index compared to their respective obese controls. This effect of *A. boonei* extract was similar to the result reported earlier by Onyeneke and Anyanwu²³, although it was of a high fat diet. Some plants contain saponins, polyphenols and flavonoids which have pancreatic lipase inhibitory effect³²; it inhibits these lipases from hydrolyzing the ingested fat into absorbable free fatty acids and monoglycerides. The decreased absorption of ingested fat leads to an overall decreased caloric absorption, in turn leading to weight loss³³.

It is well known that excess sugar in the human diet can be converted both into glycerol and fatty acids and, thus, into lipids such as triglycerides³⁴. This explains why the triglyceride, total cholesterol, VLDL cholesterol and LDL cholesterol levels in the HCD obese rats were significantly higher than the normal control (Table 3).

The HCD obese rats had significant increase in glucose and insulin concentration. This effect is similar to that found in HFD obese rats as indicated by Zhang *et al.*³⁵ and Amin and Nagy²⁵. The *A. boonei* extract significantly decreased leptin levels. Leptin is one of the hormones that regulate hunger and satiety. Leptin is a powerful appetite suppressant, which help to regulate cravings for sweet foods³⁶. Leptin levels are excessively high in obese people and this leptin resistance is associated with weight gain. In this study, *A. boonei* extract decreased leptin level which might suggest that the plant possess appetite suppressant ability or ability to boost leptin sensitivity. Its low level will decrease quest for food by the animals and may have been responsible for the low food intake observed by the animals treated with the plant extract.

Leptin is a hormone produced mainly by adipocytes, thus leptin concentration is related to the amount and distribution of body fat³⁷ such that the heavier the body weight the higher the leptin concentration in human and rodents³⁸. Therefore, the decreased leptin level might be due to decreased fat deposits in the treated rats which might be consequent upon some bioactive components in the *A. boonei* extract.

The photomicrograph of HCD obese control showed presence of numerous prominent fat deposits in the livers and this appeared reduced in group 3. There is increasing evidence that non-alcoholic fatty liver disease often represents a component of the metabolic syndrome characterized by obesity, hyperinsulinemia, peripheral insulin resistance, diabetes, hypertriglyceridemia and hypertension^{39,40}.

Fat deposits were not present in the heart of HCD obese control, although there were signs of myocardial infarction which was not much different with that of the treated groups. Abnormal blood lipid levels, that is high total cholesterol, high levels of triglycerides, high levels of low-density lipoprotein (LDL, "bad" cholesterol) or low levels of high-density lipoprotein (HDL, "good" cholesterol) cholesterol all increase the risk of heart disease like myocardial infarction and stroke⁴¹. High total cholesterol levels, which may be caused by many factors, are associated with an increased risk of cardiovascular disease^{42,43}.

Conclusion

The high carbohydrate diet formulated for this study induced obesity in the male Wistar rats and gave rise to increased food intake, body weight, fat mass, cholesterol levels, glucose levels, insulin and leptin levels; this may have implications for the progress of obesity related problems. The ethanolic extract of *A. boonei* stem bark which reduced the food intake, body weight, fat mass and leptin levels might possess anti-obesity potential.

References

1. WHO (2009). Obesity and Overweight. Organization. Retrieved February 22, 2009. www.who.int/dietphysicalactivity/media/en/gsf Obesity.pdf
2. Pagotto U., Vanuzzo D., Vicennati V., Pasquali R.G. (2008). Pharmacological therapy of obesity. *G. Ital. Cardiol. (Rome)*, 9:83–93.
3. Marović D. (2008). Elevated body mass index fatty liver. *Srp. Arh. Celok. Lek.*, 136:122–5.
4. Lavie C.J., Artham S.M., Milani R.V., Ventura H.O. (2008). The obesity paradox: impact of obesity on the prevalence prognosis of cardiovascular diseases. *Postgraduate Medicine*, 120:34–41.
5. Albrecht, C., Baynes, K. and Sardini, A. (2004). Two novel missense mutations in ABCA1 result in altered trafficking and cause severe autosomal recessive HDL deficiency. *Biochemical Biophysical Acta.*, 89: 7–57.
6. WHO (2000). World Health Organization: Part I: The problem of overweight and obesity: In World Health Organization. Obesity: preventing and managing the global epidemic. Geneva: WHO Technical Report Series; 2000.
7. Yach, D., Stuckler, D. and Brownell, K. D. (2006). Epidemiologic and economic consequences of the global epidemics of obesity and diabetes. *Natural Medicine*, 12 (1): 62–6.
8. Slyper, A. H. (2004). The pediatric obesity epidemic: causes and controversies. *J. Clin. Endocrinology Metabolism*, 89 (6):2540-7.
9. Terashima, H. (2003). Names, use and attributes of plants and animals among the Ituri forest foragers: a comparative ethnobotanical and ethnozoological study. *African Study Monographs*, 28 (Suppl.):7–24.
10. Abel, C. and Busia, K. (2005). *Alternative Medicine Review*, 2005. An Exploratory Ethnobotanical Study of the Practice of Herbal Medicine by the Akan Peoples of Ghana (Ethnobotanical Study: Herbal Medicine in Ghana), pp.42.
11. Treatise, G. E. and W. C. Evans (1989). *Pharmacognosy*. 11th Edn. Brailliar Tiridel Can. Macmillan Publishers. Ltd, Ibadan, 289.
12. Sofowora (1993). *A Medicinal Plants and Traditional Medicine in Africa*. Spectrum Books Ltd, Ibadan, pp. 289.
13. Shah, M. A., Garg, S. K. and Garg, K. M. (1997). Subacute toxicity studies on Pendimethalin in rats. *International Journal of Pharmacology*, 29: 322-324.
14. Burger, C., Fischer, D. R., Cordenunzi, D.A., Batschauer de Borja, A. P., Filho V.C. and A. R., Soares dos Santos, 2005. Acute and subacute toxicity of the hydroalcoholic extract from *Wedelia paludosa* (*Acmela brasiliensis*) (Asteraceae) in mice. *Journal of Pharmacology Science* (www.cspCanada.org), 8: 370-373.
15. Taylor, B.A. and Phillips, S.J. (1996). Detection of obesity QTLs on mouse chromosomes 1 and 7 by selective DNA pooling. *Genomics*, 34(3):389-398.
16. Trinder, P. (1969). Determination of glucose in blood using glucose oxidase with an alternative oxygen acceptor. *Annals of Clinical Biochemistry*, 6: 24-27.
17. Tietz, N. W., (1990). *Clinical Guide to Laboratory Tests*, Second Edition W. B. Saunders Company, Philadelphia, USA, pp. 554-556.

18. Lopes-Virella, M.F., Stone, P., Ellis, S. and Colwell, J.A. (1977). Cholesterol determination in HDL separated by three different method. Clin. Chem. 23: 882 – 884.
19. Friedewald, W. T., Levy, R. I. and Fredrickson, D. S. (1972). Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without the use of preparative ultracentrifuge. Clinical Chemistry, 18:499-502.
20. Dahiru, D. and Obidoa, O. (2008). Evaluation of the antioxidant effects of *Zizyphus mauritiana* lam. Leaf extracts against chronic ethanol induced hepatotoxicity in rat liver. Afr. J. Trad. CAM. 5(1): 39-45.
21. Akinmoladun, A. C., Ibukun, E. O., Afor, E., Akinrinola, B. L., Onibon, T. R., Akinboboye, A. O., Obuotor, E. M. and Farombi, E. O. (2007). Chemical constituents and antioxidant activity of *Alstonia boonei*. African Journal of Biotechnology, 6 (10): 1197-1201.
22. Amole, O. O. and Ilori, O. O. (2010). Antimicrobial activity of the aqueous and ethanolic extracts of the stem bark of *Alstonia boonei* International Journal of Phytopharmacology, 1(2):119-123.
23. Onyeneke, E. C and Anyanwu, G. O (2013). Effect of *Alstonia boonei* De Wild on High Fat Diet Induced Obesity in Male Wistar Rats. European Journal of Scientific Research, 112 (3): 292-299.
24. Hofmann, S. M. and Tschöp, M. H. (2009). Dietary sugars: a fat difference. Journal of Clinical Investigation, 119, 5, 1089–1092.
25. Amin, K. A. and Nagy, M. A. (2009). Effect of Carnitine and herbal mixture extract on obesity induced by high fat diet in rats. *Diabetol. Metab. Syndr.*, 1: 17-24.
26. Haslam, D. W. and James, W. P. (2005). Obesity. Lancet, 366 (9492): 1197–209.
27. Madsen, A. N., Hansen, G., Paulsen, S. J., Lykkegaard, K., Tang-Christensen, M., Hansen, H. S., Levin, B. E., Larsen, P. J., Knudsen, L. B., Fosgerau, K. and Vrang, N. (2010). Long-term characterization of the diet-induced obese and diet-resistant rat model: a polygenetic rat model mimicking the human obesity syndrome. Journal of Endocrinology, 206: 287–296.
28. Mistry, K. G., Deshpande, S. S., Shah, G. B. and P. V. Gohil. (2011). Effect of Sarpogrelate in high fat diet induced obesity in rats. Asian Journal of Pharmacology and Biology Research, 1(4): 441-446.
29. Fernandez, C. D. B., Bellentani, F. F., Fernandes, G. S. A., Perobelli, J. E., Favareto, A. P. A., Nascimento, A. F., Cicogna, A. C. and Kempinas, W. D. G. (2011). Diet-induced obesity in rats leads to a decrease in sperm motility. Reproduction Biology and Endocrinology, 9:32.
30. Raji, Y., Salman, T. M., and Akinsomiso, O. S. (2005). Reproductive functions in male rats treated with methanolic extract of *Alstonia boonei* stem bark. African Journal of Biomedical Research, 8: 105 – 111.
31. Jeyakumar, S. M., Lopamudra, P., Padmini, S., Balakrishna, N., Giridharan, N. V., Vajreswari, A. (2009). Fatty acid desaturation index correlates with body mass and adiposity indices of obesity in Wistar NIN obese mutant rats strains WNIN/Ob and WNIN/GR-Ob. Nutrition Metabolism, 6:27.
32. Han, L. K., Zheng, Y. N., Yoshikawa, M., Okuda, H. and Kimura, Y. (2005). Anti-Obesity Effects of Chikusetsusaponins Isolated from *Panax japonicus* Rhizome. BMC Complementary and Alternative Medicine, 5(9):1-10.
33. Tsujita, T., Takaichi, H., Takaku, T., Aoyama, S. and Hiraki, J. (2006). Antiobesity action of E-Polylysine, a potent inhibitor of pancreatic lipase. Journal of Lipid Research, 47 (8): 1852-1858.
34. Murray, R. K., Granner, D. K., Mayes, P. A., and Rodwell, W. V. (2003). Harper's Illustrated Biochemistry, Twenty-Sixth Edition, McGraw-Hill Companies, USA, pp. 1-129.
35. Zhang, M., Lv, X. Y., Li, J., Xu, Z. G. and Chen, L. (2008). The characterization of high-fat diet in multiple low-dose streptozotocin induced type 2 diabetes rat model. Experimental Diabetes Research; 2008:704045.
36. Ninomiya, Y., Shigemura, N., Yasumatsu, K., Ohta, R., Sugimoto, K., Nakashima, K. and Lindemann, B. (2002). Leptin and sweet taste. Vitamins and Hormones., 64:221-48.
37. Mayes, J. S. and Watson, G. H. (2004). Direct effects of sex steroid hormones on adipose tissues and obesity. Obesity Reviews, 5(4):197-216.
38. Aizawa-Abe, M., Ogawa, Y., Masuzaki, H., Ebinara, K., Satoh, N., Iwai, H., Matsuoka, N., Hayashi, T., Hosoda, K., Inoue, G., Yoshimura, Y. and Nakao, K. (2000). Pathophysiological role of leptin in obesity-related hypertension. Journal of Clinical Investigation, 105(9):1243-1252.
39. Cortez-Pinto, H., Camilo, M. E., Baptista, A., De Oliveira, A. G. and De Moura, M. C. (1999). Non-alcoholic fatty liver: another feature of the metabolic syndrome? Clinical Nutrition, 18, 353–358.
40. Sharabi, Y. and Eldad, A. (2000). Nonalcoholic fatty liver disease is associated with hyperlipidemia and obesity. American Journal of Medicine, 109:171.
41. Clarke, R., Frost, C., Collins, R., Appleby, P. and Peto, R. (1997). Dietary lipids and blood cholesterol: quantitative meta-analysis of metabolic ward studies. British Medical Journal (Clinical research ed.) 314 (7074): 112–7.
42. Bucher, H. C., Griffith, L. E. and Guyatt, G. H. (1999). "Systematic review on the risk and benefit of different cholesterol-lowering interventions." Arteriosclerosis Thrombosis Vascular Biology, 19 (2): 187–195.
43. Lewington, S., Whitlock, G., Clarke, R., Sherliker, P., Emberson, J., Halsey, J., Qizilbash, N., Peto, R. and Collins, R. (2007). "Blood cholesterol and vascular mortality by age, sex, and blood pressure: a meta-analysis of individual data from 61 prospective studies with 55,000 vascular deaths". Lancet 370 (9602): 1829–39.