BRC 2001077/14309

# Fungi present in the root zone of Amaranthus hybridus

## G. P. Oyeyiola

Department of Biological Sciences, University of Ilorin, Ilorin, Nigeria

(Received May 8, 2001)

ABSTRACT: Fungi present in the root zone of Amaranthus hybridus were isolated and identified. The fungi were Aspergillus niger, Aspergillus terreus, Aspergillus versicolor, Rhizopus stolonifer, Mucor pusillus, Cladosporium sp.' Alternaria alternata, Aspergillus fumigatus, Penicillium oxalicum, Chaetomium sp., Penicillium citrinum and Rhizopus oligosporus. R. stolonifer, A. niger, A. terreus and P. oxalicum predominated in both the rhizosphere soil and the rhizoplane. Aspergillus versicolor, Alternaria altenata and Aspergillus fumigatus were present in the root zone but were absent from the non-rhizopsphere soil. The experimental soil was sandy loam in texture. Rhizosphere effect was well pronounced and it increased progressively with increase in plant age until the tenth week after sowing and then declined.

Key Words: Root zone fungi; Amaranthus hybridus; Rhizosphere effect.

#### Introduction

The root zone (1) is an area of intense microbial activity (2). This is because microorganisms depend on easily available organic compounds for survival. Such compounds occur in the root zone in the form of root exudates and sloughed off root cells (3,4). Microorganisms in the root zone are not passive. They may influence both positively or negatively some aspects of plant growth like root morphology, root-to-shoot weight ratios, uptake of calcium and rubidium, uptake of phosphorus and sulphur, rate of development, onset of flowering and crop yield (2). Microorganisms present in the root zone vary with soil types and plant species (2,5,6) because different plant species produce different effects upon the microorganisms (2,7-9). Fungi represent an important component of the soil microbial community (10). Fungi in the root zone may play a role in the nutrition and well being of the plant or contribute to the plant's resistance to soil borne pathogens (11,12).

Reports are available on the root zone mycoflora of a number of plants (6,11,13-16). However, scanty information is available on the root zone mycoflora of *Amaranthus hybridus* which is a vegetable consumed by millions of Nigerian. The present study is on the root zone mycoflora of this plant from ecological point of view and in comparison with the non-rhizosphere soil.

#### **Materials and Methods**

Texture, organic matter content, pH, moisture content and water holding capacity of the soil were determined prior to sowing of seeds. The physical and chemical characteristics were determined by the method of Pramer and Schmidt (17) and that of Olaitan and Lombin (18).

Rhizosphere soil, non-rhizosphere soil and plant root samples were collected and serial dilutions were prepared from them as previously described (11, 16, 19, 20), using fresh samples in each case. Isolation of fungi was made from desired dilutions and the plates were incubated at 30°C. Potato dextrose agar (PDA) and malt extract agar (MEA) were the culture media used and to each medium was added appropriate amounts of streptomycin and rose Bengal (21). Colony counts and subculturing were carried out after 72 hours. Representative colonies were taken out for identification and stock cultures were prepared on PDA slants in McCartney bottles and preserved in the refrigerator. reference was made to appropriate literature (22,23) to get the names of the isolates.

The ratio of the number of fungi in the rhizosphere soil and non-rhizosphere soil (R/S ratio) was calculated using the formula:

No. of fungi/g of rhizosphere soil (R) No. of fungi/g of non-rhizosphere soil (S)

#### **Results and Discussion**

The experimental soil was sandy loam (Table 1). Sandy loam has been used for the cultivation of plants by some previous workers (16, 24). Greater variety of fungi occurred in the rhizosphere than in the rhizoplane (Table 2). This could be attributed to the fact that the roots are the source of exudations. These exudations have selective effect on microorganisms in the vicinity of roots (2). This effect will be most marked at the root-soil interface (25, 26) reducing in intensity with increasing distance from the root surface. Therefore, less check will be exercised on fungi growing some distance away from the roots, thereby allowing a greater range of fungi to develop than on the root surface. Odunfa (6) also encountered more fungal species in the rhizosphere than in the rhizoplane of sorghum.

| Table 1: | Some physical | and chemical | characteristics | of the experimenta | l soil | (prior to | sowing o | of seeds). |
|----------|---------------|--------------|-----------------|--------------------|--------|-----------|----------|------------|
|          |               |              |                 |                    |        |           |          |            |

| Characteristics        | Values     |
|------------------------|------------|
| Water holding capacity | 0.28 ml/g  |
| рН                     | 7.4        |
| Moisture content       | 4.5%       |
| Organic matter         | 3.0%       |
| Mineral fraction:      |            |
| Sand                   | 84.10%     |
| Silt                   | 5.77%      |
| Clay                   | 10.13%     |
| Soil texture           | Sandy loam |

| Isolates                   | Period of sampling (weeks) |      |      |      |      |      |      |  |
|----------------------------|----------------------------|------|------|------|------|------|------|--|
| -                          | 2                          | 4    | 6    | 8    | 10   | 12   | 14   |  |
| From rhizosphere soil      |                            |      |      |      |      |      |      |  |
| Aspergillus niger          | 50.0                       | 30.2 | 32.3 | 34.0 | 35.1 | 30.6 | 31.0 |  |
| Aspergillus terreus        | —                          | 27.0 | 26.6 | 30.2 | 30.4 | 28.2 | 25.3 |  |
| Aspergillus versicolor     | _                          | _    | _    | _    | _    | 2.6  | 2.4  |  |
| Rhizopus stolonifer        | 38.2                       | 16.9 | 24.0 | 21.8 | 20.7 | 20.0 | 20.0 |  |
| Mucor pusillus             | _                          | _    | _    | _    | 3.0  | 2.3  | 3.5  |  |
| Cladosporium sp.           | 7.3                        | _    | _    | _    | _    | _    | _    |  |
| Penicillium oxalicum       | _                          | 20.0 | 13.0 | 10.0 | 7.6  | 14.9 | 17.4 |  |
| Aspergillus fumigatus      | 4.5                        | 2.3  | _    | _    | _    | _    | _    |  |
| Alternaria alternata       | _                          | 3.6  | 4.1  | 4.0  | 3.2  | 1.4  | 1.4  |  |
| From rhizoplane:           |                            |      |      |      |      |      |      |  |
| Rhizopus stolonifer        | 30.1                       | 28.0 | 26.6 | 27.0 | 20.2 | 19.2 | 19.1 |  |
| Aspergillus niger          | 26.0                       | 22.7 | 23.0 | 20.1 | 21.6 | 20.0 | 20.0 |  |
| Aspergillus terreus        | 20.0                       | 23.6 | 27.3 | 28.1 | 29.1 | 29.7 | 29.9 |  |
| Penicillium oxalicum       | 16.7                       | 19.4 | 19.9 | 24.8 | 25.0 | 25.8 | 26.5 |  |
| Aspergillus versicolor     | _                          | _    | _    | _    | 4.1  | 2.8  | 2.5  |  |
| Alternaria alternata       | _                          | _    | _    | _    | _    | 2.5  | 2.0  |  |
| Aspergillus fumigatus      | 7.2                        | 6.3  | 3.2  | _    | _    | _    | _    |  |
| From non-rhizosphere soil: |                            |      |      |      |      |      |      |  |
| Aspergillus niger          | 23.1                       | 23.9 | 36.0 | 61.3 | 62.0 | 50.3 | 44.5 |  |
| Aspergillus terreus        | 30.0                       | 25.1 | 23.8 | _    | _    | _    | _    |  |
| Penicillium oxalicum       | 26.2                       | 27.0 | _    | _    | _    | _    | _    |  |
| Chaetomium sp.             | 20.7                       | 24.0 | 40.2 | 38.7 | 38.0 | 39.6 | 40.1 |  |
| Penicillium citrinum       | _                          | _    | -    | -    | -    | 10.1 | 15.4 |  |

Table 2: Frequency of occurrence of fungi in the rhizosphere soil and rhizoplane of *Amaranthus hybridus* and the non-rhizosphere soil.

The number of fungi in the rhizosphere and rhizoplane increased as the plants aged, reaching a maximum at the  $10^{th}$  week and subsequently declining (Fig. 1). Rovira (2) reported that generally the number of organisms in the rhizosphere and the rhizoplane increased with age of the plant. It was found in this study that the fungi present in the rhizosphere differed both quantitatively and qualitatively from those present in the rhizoplane, throughout the period of plant growth. Similar pattern had been reported by previous workers (6, 11, 26). Brown (27) also submitted that fungal populations differ in the two areas of the root region and change as plants mature.

Throughout the period of this study, more fungi occurred in the rhizosphere than in the non-rhizosphere quantitatively (Fig. 1). This is also reflected in the R/S ratios obtained (Fig. 2). Bowen and Rovira (12) reported that microbial growth is much more stimulated in the rhizosphere than in soil free of palnt roots, a

phenomenon called the rhizosphere effect. Such increase in the number of fungi in the rhizosphere over those in the non-rhizosphere has been observed by previous workers (6, 24, 28).

The number of fungi in the rhizosphere increased progressively as the plants aged until the  $10^{th}$  week, and subsequently declined (Fig. 1). Similar progressive increases in microbial populations in the rhizosphere of plants up to a particular stage of growth, with a subsequent fall thereafter have been reported by some workers (2, 3, 28, 29).

### References

- 1. Oyeyiola, G. P. and Hussain, H. S. N. (1992) Fungal populations in the rhizosphere and rhizoplane of wheat grown in Kadawa in Kano area of Northern Nigeria. Biosci. Res. Commun. 4(2), 129 133.
- Rovira, A. D. (1965) Interactions between plant roots and soil microorganisms. Ann. Rev. Microbiol. 19, 241 266.
- 3. Oyeyiola, G. P. (1992) Rhizosphere microbiology of wheat (*Triticum aestivum* Linn.) in the Kano area. Ph.D. Thesis, Bayero University, Kano, Nigeria.
- 4. Rouatt, J. W.; Peterson, E. A.; Katznelson, H. and Henderson, V. E. (1963) Microorganisms in the root zone in relation to temperature. Can. J. Microbiol. 9, 227 236.
- 5. Peterson, E. A. 91961) Observations on the influence of plant illumination on the fungal flora of roots. Can. J. Microbiol. 7, 1 6.
- Odunfa, V. S. A. (1979) The rhizosphere mycoflora of sorghum (Sorghum bicolour L. Moench). Nig. J. Sci. 13, 363 – 370.
- 7. Lochhead, A. C. (1952) Soil Microbiology. Ann. Rev. Microbiol. 6, 185 206.
- 8. Imam, S. and Nusrath, M. (1987) Varietal variation in the rhizosphere and rhizoplane mycoflora of *Cajanus cajan* (L.) Millsp. with special reference to wilt disease. Indian J. Bot. 10, 126 129.
- Liljeroth, E. and Baath, E. (1988) Bacteria and fungi in roots of different barley varieties (*Hordeum vulgare* L.) Biol. Fertil. Soil 7, 53 – 57.
- 10. Kaufman, D. D.; Williams, L. E. and Summer, C. B. (1963) Effect of planting medium and incumation temperature on growth of fungi in soil-dilution plates. Can. J. Microbiol. 9, 741 751.
- 11. Odunfa, V. S. A. and Oso, B. A. (1979) Fungal population in the rhizosphere and rhizoplane of cowpea. Trans. Brit. Mycol. Soc. 73, 21 26.
- 12. Bowen, G. D. and Rovira, A. D. 91976) Microbial colonization of plant root. Ann. Rev. Phytopathol. 14, 121 144.
- McDonald, D. (1968) Soil fungi and fruit of the groundnut (*Arachs hypogeae*). Samaru Miscellaneous Papers No. 28. Institute of Agricultural Research, Ahmadu Bello University, Zaria, Nigeria.
- McDonald, D. (1968) Influence of the developing groundnut fruit on soil mycoflora in Nigeria. Trans. Brit. Mycol. Soc. 53, 393 – 406.
- 15. Odunfa, V. S. A. (1980) Fusaria associated with the roots of cowpea. Nig. J. Agric. Sci. 2, 53 58.
- Akinyanju, J. A. and Fadayomi, O. (1989) Effect of Diuron on sugarcane rhizosphere microbial populations. Nig. J. Bot. 2, 49 – 58.
- 17. Pramer, D. E. and Schmidt, E. L. (1964) Experimental Soil Microbiology. Burges Publishing Company. Minneapolis, Minnesota, U.S.A.
- Olaitan, S. O. and Lombin, G. (1984) Introduction to Tropical Soil Science. (Onazi, O. C., ed.). Macmillan Publishers Ltd., London.
- 19. Oyeyiola, G. P. and Hussain, H. S. N. (1991) Bacterial and actinomycete populations in the rhizosphere and rhizoplane of wheat grown at Kadawa in Kano area of Northern Nigeria. Biosci. Res. Commun. 3, 119 130.
- Oyeyiola, G. P. and Hussain, H. S. N. (1992) Rhizosphere and rhizoplane microorganisms of wheat grown at Kura area of Kano, Northern Nigeria. Proceedings of the 32<sup>nd</sup> Annual Conference of Science Association of Nigeria, University of Ilorin, Ilorin, Nigeria, 17(2), 44 – 58.
- Doran, J. W. (1980) Microbial changes associated with residue management with reduced tillage. Soil Science Society of America Journal 44, 518 – 524.
- 22. Onions, A. H. S.; Allsopp, D. and Eggins, H. O. W. (1981) Smith's Introduction to Industrial Mycology. 7<sup>th</sup> Edition. Edward Arnold Publishers Ltd., London.
- 23. Samson, R. A. and van Reenen-Hoekstra, E. S. (1988) Introduction to food-borne fungi. 3<sup>rd</sup> Edition. Centraabureau voor Schimmecultures, Netherlands.
- 24. Peterson, E. A, (1958) Observations on fungi associated with plant roots. Can. J. Microbiol. 4, 257 265.
- 25. Cook, F. D. and Lochhead, A. G. (1959) Growth factor relationships of soil microorganisms as affected by proximity to the plant root. Can. J. Microbiol. 5, 323 334.
- Abdel-Hafez, S. I. I. (1982) Rhizosphere and rhizoplane fungi of *Triticum vulgare* cultivated in Saudi Arabia. Mycopathologia 78, 79 – 86.

- 27. Brown, M. E. (1975) Rhizoplane microorganisms opportunists, bandits or benefactors. In: Soil Microbiology. (Walker, N. ed.). Butetrworths, London. pp. 21 – 24.
- Abdel-Rahim, A. M.; Baghadami, A. M. and Abdalla, M. H. (1983) Studies on the fungus flora in the rhizosphere of sugar cane plants. Mycopathologia 81, 183 186.
  Rouatt, J. W. (1959) Initiation of the rhizosphere effect. Can. J. Microbiol. 5, 67 71.